Fast Diffusion of Long Guest Rods in a Lamellar Phase of Short Host Particles

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We investigate the dynamic behavior of long guest rodlike particles immersed in liquid crystalline phases formed by shorter host rods, tracking both guest and host particles by fluorescence microscopy. Counterintuitively, we evidence that long rods diffuse faster than short rods forming the one-dimensional ordered smectic-A phase. This results from the larger and noncommensurate size of the guest particles as compared to the wavelength of the energy landscape set by the lamellar stack of liquid slabs. The long guest particles are also shown to be still mobile in the crystalline smectic-B phase, as they generate their own voids in the adjacent layers.

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A crowded environment usually causes a slowing down of the dynamics of the constituting particles [1–3]. In the macroscopic world, traffic jams represent a good illustration of frustrated motion due to crowding, while at microscopic scales frustrated motion is observed, for instance, in polymeric [4,5] and colloidal glasses [6,7]. Generally, the degree of frustration strongly depends on the size ratio between the frustrated guest particle and the host particles that cause the frustration. When this ratio is very high then the crowding is experienced by an effective viscosity and the Stokes-Einstein relation is obeyed [8]. In the opposite case where the guest particles are much smaller, e.g., beads in a filament network, the mesh size of that network sets the degree of frustration [9–11]. As a general rule reported in the literature, large guest particles are slower than small host particles and small guest particles are faster than large host particles. In this Letter, we show that this rule does not always hold as large guest particles can be more mobile than the small host particles, depending on the self-organization of the latter.

In most studies on crowding, the motion of tracers is tracked in amorphous media [8–14]. The effect of crowding has also been shown in highly ordered phases, in which the single particle dynamics is strongly affected [15,16]. For instance, in three-dimensional colloidal crystals, the dynamics is determined by the local mobility at the crystal lattice points and by the existence of vacancies [17,18]. Reducing the dimensionality of the positional ordering from 3D to 1D leads to an unexpected hopping-type diffusion [19-22]. This has been first observed in the model system of filamentous viruses [23,24] organized in a smectic-A phase, which can be considered as a one-dimensional stack of liquid slabs [25,26]. Quasiquantized steps matching the smectic layer spacing L_{layer} have been evidenced using single particle tracking by fluorescent microscopy [27]. Here, the particles essentially behave as Brownian particles in a onedimensional potential [19,27] as the jumping rates depend on the ordering potential associated with the lamellar organization. In biological membranes, which can be seen as single smectic slabs [28,29], different macromolecules such as proteins are transported through the membranes, usually formed by lipids [30]. This leads to the question of *how transport of guest particles, which are not part of the structure forming materials, occurs through highly ordered states*. In most studies where the diffusion of guest particles through a structured background is investigated, the guest particles are smaller than the typical length scale that characterizes the host system, both in simulations [31,32] and in experiments [33–37]. Here, we address the opposite limit from an experimental point of view: How is the mobility of a guest particle affected by a surrounding energy landscape that has a smaller length scale than the guest particle?

For this study, we have devised a model system by introducing a tracer amount of long, noncommensurate, guest rods in a host smectic phase composed of shorter host rods, as schematically represented in Fig. 1(a). Having guest particles that are longer than the host layer spacing for which $L_{\text{laver}} \cong L_{\text{host}}$ [26], implies that the guest rods have to be accommodated in more than one smectic layer, exceeding therefore the typical length scale of the host ordering potential. Filamentous viruses are ideally suited for such fundamental studies, as they are intrinsically monodisperse but can be produced in different lengths [25]. Furthermore, their colloidal size allows for their imaging and tracking at the single particle level. These charged rodlike particles have been shown to behave nearly as hard rods, by defining an effective diameter accounting for their electrostatic repulsion at different ionic strengths [26]. We will show that the diffusion of the noncommensurate particles through the one-dimensional ordered smectic-A (SmA) and through the crystalline smectic-B (SmB) phases formed by shorter host rods is less sensitive to the smectic potential and that they permeate the layers faster because they are longer.

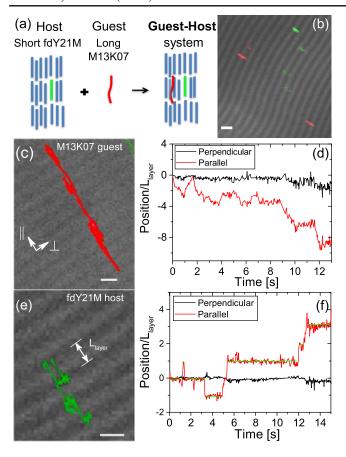


FIG. 1. (a) Schematic representation of the guest-host system exhibiting a smectic organization: the host is formed by short fdY21M viruses with both tracer amounts of labeled host particles (green, fdY21M) and of long labeled semiflexible guest rods (red, M13K07), which are 1.3 times longer than the host ones. (b) Overlay of a differential interference contrast microscopy image evidencing smectic layers, and of a fluorescence image, displaying the dual labeling of the host and guest particles. (c) Overlay of a differential interference contrast picture and a M13K07 guest trajectory for which rapid diffusion is observed through the layers. (d) Corresponding displacements of the M13K07 rod, parallel (red) and perpendicular (black) to the normal of the smectic layer. (e) Example of trace for a fdY21M host particle in the lamellar phase of layer spacing L_{layer} . (f) Associated displacement where a jumping-type diffusion process is clearly evidenced. The green lines are obtained by the jump recognition algorithm and define the residence time τ_{res} that rods spend within a smectic layer before a hopping-type event. The scale bars represent 1 μ m.

We used two mutants of the filamentous bacteriophage fd to create our guest-host model system with fdY21M virus as a short stiff host (contour length $L_{\rm fdY21M}=0.91\,\mu{\rm m}$, persistence length $P_{\rm fdY21M}=9.9\,\mu{\rm m}$, diameter $d=7\,{\rm nm}$) and M13K07 helper phage as a long guest semiflexible rod ($L_{\rm M13K07}=1.2\,\mu{\rm m}$, $P_{\rm M13K07}=2.8\,\mu{\rm m}$, $d=7\,{\rm nm}$) [27,38,39], both prepared following standard biological protocols [40]. Consequently, the guest-host length ratio is $L_{\rm M13K07}/L_{\rm fdY21M}\cong L_{\rm M13K07}/L_{\rm layer}=1.3$, as shown in Fig. 1(b). The choice of fdY21M as stiff rods for the host

matrix enables us to significantly increase the smectic-A range compared to semiflexible fd-wt suspensions [26], allowing therefore for a more accurate study of the dynamics close to both the chiral nematic-SmA and SmA-SmB phase transitions. A second batch of fdY21M viruses was prepared, for which an extended amplification time of the bacteria cultures was applied, yielding the formation of a low fraction of multimeric viruses in the suspension. The virus contour length being fixed by the size of its DNA, a few percent of dimers and trimers (containing, respectively, 2 and 3 times the virus genome) exist in this polydisperse batch (see the Supplemental Material for details [41]). We labeled fdY21M and M13K07 with green and red fluorescent dyes, respectively, to easily distinguish them by fluorescence microscopy. Labeled particles were added in a ratio of one labeled particle over 10⁵ nonlabeled particles such that trajectories of individual rods could be recorded (Fig. 1). The amount of labeled particles is small enough to assume that the phase behavior of the host system remains unaffected. The multimeric batch containing fdY21M dimers and trimers in a small amount exhibits similar quantitative phase behavior as the batch of pure single host particles (see below). A broad range of concentrations of aqueous suspensions of viruses (in TRIS-HCl-NaCl buffer, pH 8.2, ionic strength 20 mM) has been investigated from the (chiral) nematic to the smectic-A and smectic-B phases [26].

Figures 1(c)–1(f) show two typical trajectories recorded in the smectic-A phase for both M13K07 guest and fdY21M host particles. Qualitatively, two main features distinguish their respective dynamic behavior: (1) the long guests do not exhibit hopping-type events as sharp as those observed for the host particles, which also stay longer in a given smectic layer having therefore a longer residence time $\tau_{\rm res}$; (2) the parallel displacement (along the director or, equivalently, along the normal of the smectic layers indicated by the z axis) for a given observation time is bigger for the long M13K07 guest viruses. We quantify these observations by comparing the ordering potentials that are effectively felt by both tracer particles and by measuring the associated dynamic behavior of the guest and host particles through their mean square displacements (MSDs).

The 1D ordering potential $U_{\rm layer}$, which characterizes the lamellar organization of the smectic phase, can be obtained experimentally from the particle fluctuations with respect to the middle of the layer [19]. The distribution of these fluctuations is proportional to the probability P(z) of finding the center of mass of the rod-shaped particles with respect to the middle of the layer, via the Boltzmann factor $P(z) \sim e^{-U_{\rm layer}(z)/k_BT}$ [27]. Figure 2(a) displays the concentration dependence of the potential barrier U_0 for the different particles. While for the short host particles the onset of $U_{\rm layer}^{\rm fdY21M}$ occurs at the N-SmA transition, as expected, the onset of $U_{\rm layer}^{\rm M13K07}$ for the long guests is shifted to much higher concentrations. This means that M13K07 guest particles do not feel the underlying periodic potential

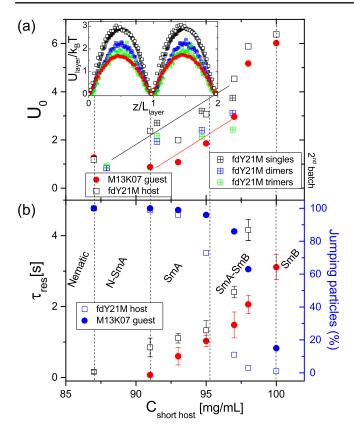


FIG. 2. Concentration dependence of (a) smectic potential barriers obtained after deconvolution from the point-spread function of the microscope for long noncommensurate guest (red solid circles), long commensurate guest (blue and green crossed squares), and host (black open squares and crossed squares) particles. The solid lines are guides to the eye. Inset: raw data (before deconvolution) of the energy landscape at a host concentration of $C_{\rm fdY21M}=95~{\rm mg/mL}$. (b) Average residence time $\tau_{\rm res}$ for which the particles remain in a layer before a hopping-type event, and percentage of jumping particles during the acquisition time window of 15 s (blue symbols). The vertical dashed lines indicate the phase transitions.

as strongly as the fdY21M host ones. Despite their commensurability with the layer spacing, multimeric fdY21M viruses (dimers and trimers) exhibit lower potentials than singles, but still higher potentials than M13K07 particles. This can be understood by considering that the dimer center of mass experiences a maximum of the potential, contrary to singles, which are trapped in a minimum. Such an explanation can be extended to trimers, whose particle body feels two maxima of the smectic potential, reducing accordingly the resulting potential felt by these multimeric viruses. The difference in potential between guest and host particles persists at the SmA-SmB transition where $U_{\mathrm{layer}}^{\mathrm{M13K07}}$ continues to increase, while for the short hosts the diffusion freezes in the crystalline smectic-B phase. This corresponds to a divergence of the ordering potential, which impedes its accurate determination. It is worth mentioning that both particles have the same concentration dependence, since the slope is similar for both curves. This is directly reflected in the ratio of particles that jump layers over particles that do not jump (within the time window of a trajectory) [Fig. 2(b) in blue]: hopping-type events can still be found in the SmB phase for the long M13K07 guests at concentrations where the host particles ceased to jump. Similarly, the average residence time τ_{res} , which is calculated from the histogram of residence times for all particles, strongly increases around the SmA-SmB transition for the host particles, while $\tau_{\rm res}$ rises more smoothly throughout that transition for the long guest ones, as shown in Fig. 2(b). Note that for the longest residence time $\tau_{\rm res} \simeq 4$ s, only 3% of the particles continue to display jumps within the observation time window of 15 s. This confirms that our acquisition time window is long enough to capture, on average, any hopping-type events occurring in the system.

Beyond the ordering potentials, the self-diffusion of the system can be quantified through the full range of concentrations by the mean square displacement (MSD), which is determined from the particle trajectories $\mathbf{r}(t)$. In complex systems, the time evolution of the MSD is not always linear as for diffusive particles, but it is more generally described by a power law:

$$MSD_{\parallel,\perp} = \langle r_{\parallel,\perp}^2(t) \rangle = 2 \times D_{\parallel,\perp} t^{\gamma_{\parallel,\perp}}, \tag{1}$$

where $D_{\parallel,\perp}$ and $\gamma_{\parallel,\perp}$ are the diffusion rates and exponents parallel (\parallel) and perpendicular (\perp) to the director, respectively. The anisotropic MSDs of the guest particles are displayed in Fig. 3 for the highest concentrations corresponding mainly to the smectic organization. Diffusion rates and exponents have been obtained from a fit of these data according to Eq. (1).

Diffusive behavior is obtained for both rod systems and in both directions in the entire nematic range. We do observe that the diffusion rates of the long guest is lower compared to the short host, especially for $D_{\perp}^{\text{M13K07}}$ in the perpendicular direction, as shown in Fig. 4, even when renormalizing by the anisotropic diffusion coefficients at infinite dilution, which partially accounts for some rod size effects (see the Supplemental Material [41]). Thus, in the nematic phase, long rods are more hindered, as one would have expected. Inversely, after the N-Sm transition, D_{\perp} is similar for both rods, while $D_{\parallel}^{\rm fdY21M}$ for the short hosts sharply decreases (Fig. 4). For fdY21M singles, dimers, and trimers, the scaling by their length of the diffusion rates works remarkably well in the smectic-A range. This confirms that the longer the commensurate particles are, the slower their diffusion is, as also directly demonstrated in the insets of Fig. 4. This is in strong contrast to the noncommensurate M13K07 guests, for which $D_{\parallel}^{\text{M13K07}}$ even displays a slight increase after the phase transition, showing that the long noncommensurate guest particles diffuse significantly faster in the smectic-A phase. The

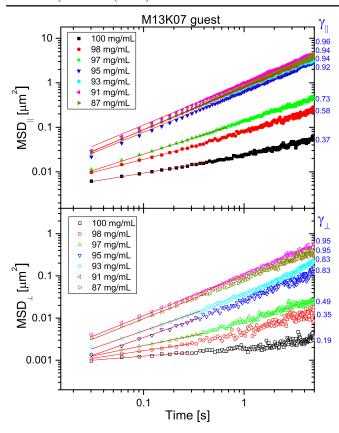


FIG. 3. Log-log representation of the mean square displacement (MSD) parallel (top) and perpendicular (bottom) to the director plotted as a function of time for the long guest M13K07 particles in the range of high host concentrations. The diffusion exponents γ obtained from the numerical fit (red lines) according to Eq. (1) are also indicated.

behavior is subdiffusive, whatever the direction, when approaching the SmB phase. It is important to stress that in this high concentration range, we observe after some time a change in the slope of the log-log MSD for all particles (Fig. 3 and the Supplemental Material [41]). The increase of the slopes, and thus of γ , is interpreted as a consequence of a void creation in the adjacent smectic layer. This results in the release of the layer constraint of the initially confined particles, allowing them to jump. The diffusion exponents γ_{\parallel} are higher for the guest particles, which is interpreted as being due to voids that are more easily generated, as compared to voids that have to be created for the host particles. This phenomenon is more apparent in the dense crystalline smectic-B phase.

The fact that no significant difference is observed between semiflexible long guests and stiff short hosts in the perpendicular diffusion is rather unexpected. Indeed, it has been shown that self-diffusion within the layers is far more pronounced for stiff rods as compared to more flexible ones of the same size [27]. The reason is that parallel and perpendicular diffusion are correlated, and that M13K07 guests feel a weaker ordering potential than the

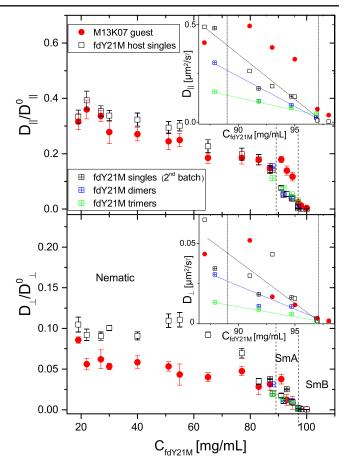


FIG. 4. Diffusion rates *normalized* by the anisotropic diffusion coefficients at infinite dilution D^0_{\parallel} and D^0_{\perp} (see the Supplemental Material [41]) for long noncommensurate guest (red solid circles), long commensurate guest (blue and green crossed squares), and host (black open squares and crossed squares) particles. Insets: enlargement of the *raw* diffusion rates in the smectic range evidencing that long (dimers and trimers) commensurate rods diffuse slower than single host ones, while long noncommensurate ones (M13K07) diffuse faster. The dotted lines are guides for the eye. The vertical dashed lines indicate the phase boundary concentrations.

constituting host particles. This accounts then for their relative promoted lateral diffusion. As a result, also, their diffusion exponents exhibit a less enhanced subdiffusivity than the host particles at the same concentration (Fig. 3 and the Supplemental Material [41]). For multimeric fdY21M particles, hindrance in the lateral diffusion increases with rod length [Fig. 4(b)], as these commensurate rods belong simultaneously to many smectic layers.

In conclusion, the general assumption that large particles always diffuse slower than small ones is not valid as soon as the length scale associated with the energy landscape formed by the host particles is smaller than the guest particle size. We proved this effect by evidencing a promoted permeation of noncommensurate long guest rods through self-assembled smectic layers of shorter host particles. This phenomenon can be understood, realizing

that the ordering potential barrier in the smectic phase is not a static value but rather the result of rod density fluctuations within the layer. A particle can therefore jump when a transient void exists in the adjacent layer. At higher concentrations, the number of voids decreases and hence the potential barrier increases. As noncommensurate long rods are always present in at least two layers at the same time, they generate their own voids facilitating their permeation. Our work should therefore stimulate further theoretical investigation for the generalization of the concept evidenced here and for the design of fast diffusers in crowded environment with potential application in cell biology and drug delivery.

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Supplementary Information:

Fast Diffusion of Long Guest Rods in a Lamellar Phase of Short Host Particles

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I. EXPERIMENTAL METHODS

We used two mutants of the filamentous bacteriophage fd to create our guest-host model system with fdY21M virus as a short stiff host (contour length $L_{fdY21M} = 0.91 \ \mu m$, persistence length $P_{fdY21M} = 9.9 \ \mu m$, diameter $d = 7 \ \text{nm}$) and M13K07 helper phage as long guest semi-flexible rod ($L_{M13K07} = 1.2 \ \mu m$, $P_{M13K07} = 2.8 \ \mu m$, $d = 7 \ \text{nm}$), both prepared following standard biological protocols [1]. Consequently, the guest-host length ratio is $L_{M13K07}/L_{fdY21M} \cong L_{M13K07}/L_{layer} = 1.3$. A second batch of fdY21M viruses was prepared, for which an extended amplification time (typically overnight instead of the usual 7 hours) of the bacteria cultures was applied, yielding to the formation of a low fraction of multimeric viruses in the suspension. The virus contour length being fixed by the size of its DNA, a few percents of dimers (14%) and trimers (2%) (containing respectively two and three times the virus genome) exist in this polydisperse batch (Fig. S1) [1].

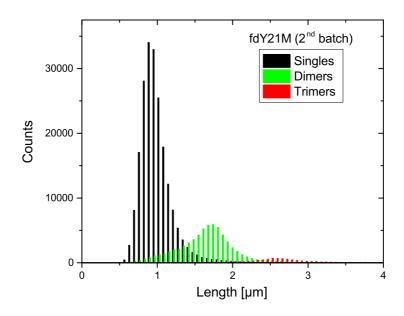


FIG. S1: Distribution of observed virus lengths obtained for fdY21M samples (2^{nd} batch containing multimeric particles after an extended incubation time of the bacteria) using particle tracking algorithms throughout the different movies. Three populations of viruses can be distinguished: singles ($L_0 \equiv L_{fdY21M} \simeq 0.9 \ \mu m$), dimers ($2L_0 \simeq 1.8 \ \mu m$), and trimers ($3L_0 \simeq 2.6 \ \mu m$) representing 84%, 14% and 2% of the total population, respectively.

We labeled both fdY21M batches with green fluorescent Alexa488-TFP (Invitrogen) and

M13K07 with the red fluorescent Dylight549-NHS ester (ThermoFischer). Green and red labeled particles were added in a ratio of one labeled particle over 10⁵ non-labeled particles such that trajectories of individual rods can be recorded (Fig. 1). The amount of labeled particles is small enough to assume that the phase behavior of the host system remains unaffected. A broad range of concentrations from the (chiral) nematic to the smectic-A and smectic-B has been investigated. Samples were prepared by introducing a droplet of aqueous suspensions of viruses (in TRIS-HCl-NaCl buffer, pH 8.2, ionic strength of 20 mM) between a cover slip and a glass slide separated by 10 μm thick mylar spacers. Samples were slightly sheared for a few seconds before sealing (UV glue NOA81) in order to obtain uniformly oriented domains. Fluorescently labeled viruses were visualized using an inverted microscope (IX-71 Olympus), equipped with a high-numerical aperture (NA) oil objective (100x PlanApo NA 1.40) and an excitation light source (X-cite series 120 Q). A dual emission image splitter (Optosplit II Andor) was used to simultaneously acquire the two fluorescent emission wavelengths on the sensor of an ultra-fast electron-multiplying camera (NEO sC-MOS Andor). The exposure time was set at 20 or 50 ms per frame depending on the virus batch, while the movie duration was limited to 15 s because of dye photobleaching and thus loss of tracked particles. A total number of about 800 trajectories per concentration was collected using a particle tracking algorithm developed initially by Crocker and Grier [2].

II. TRANSLATIONAL DIFFUSION COEFFICIENTS IN THE LIMIT OF INFINITE DILUTION

The translational diffusion coefficient D^0 of a Brownian rod in the isotropic phase is the geometrical average of the diffusion coefficients along (D_{\parallel}^0) and perpendicular (D_{\perp}^0) to the rod long axis: $D^0 = \frac{1}{3}(D_{\parallel}^0 + 2D_{\perp}^0)$. In the limit of infinite dilution, the rods move without interacting with each other and only experience hydrodynamic friction with the solvent [3]. This leads to $D_{\parallel,\perp}^0 = k_B T/\gamma_{\parallel,\perp}$ where $\gamma_{\parallel,\perp}$ are the friction coefficients. In the limit of long rods, $\gamma_{\parallel,\perp}$ are given by $\gamma_{\parallel} = \frac{2\pi\eta_0 L}{Ln\{L/d\}}$ and $\gamma_{\perp} = 2\gamma_{\parallel}$ with L and d the rod length and diameter respectively, and $\eta_0 = 10^{-3}$ Pa.s the water viscosity at room temperature [4]. Numerically, the results are provided in Table S1 for the viruses of different lengths used in this work.

Virus	$D_{\parallel}^0 \left[\mu m^2/s\right]$	$D_{\perp}^0 \ [\mu m^2/s]$
M13K07	2.76	1.38
fdY21M	3.44	1.72
Dimer (fdY21M)	1.97	0.98
Trimer (fdY21M)	1.40	0.70

TABLE S1: Translational diffusion coefficients in the limit of infinite dilution for fdY21M (single, dimer and trimer) and M13K07 rod-like viruses.

III. SELF-DYNAMICS OF THE GUEST-HOST SYSTEM

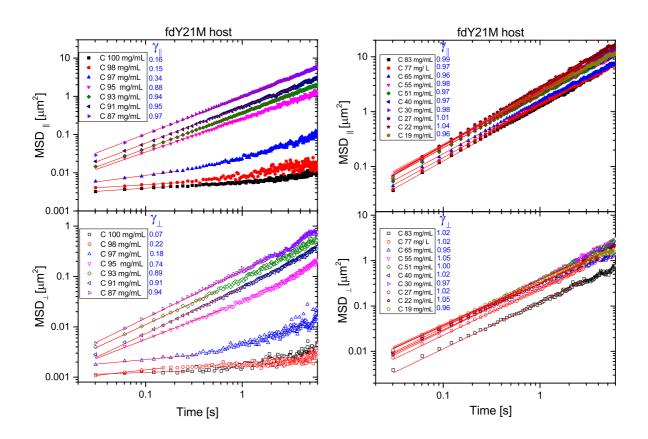


FIG. S2: Log-log representation of the parallel (top, solid symbols) and perpendicular (bottom, open symbols) mean square displacements (MSDs) for short host fdY21M particles in the smectic (left) and nematic (right) range. Blue values indicate the anisotropic exponents γ obtained from a power law fit performed according to Eq. (1) in the main text.

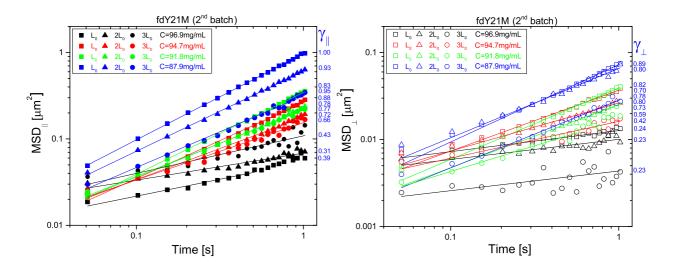


FIG. S3: Log-log representation of the parallel (left, solid symbols) and perpendicular (right, open symbols) mean square displacements (MSDs) for the second batch of fd21M particles containing multimeric viruses (L_0 singles (squares); $2L_0$ dimers (triangles); $3L_0$ trimers (circles)). Black, red, green and blue symbols refer to virus concentrations of 96.9, 94.7, 91.8 and 87.9 mg/mL, respectively. Blue values indicate the anisotropic exponents γ obtained from a power law fit performed according to Eq. (1) in the main text.

The mean square displacements (MSDs) of the guest-host system are reported in Figures 3, S2, S3 and S4. The anisotropic diffusion rates $D_{\parallel,\perp}$ and exponents $\gamma_{\parallel,\perp}$, parallel (\parallel) and perpendicular (\perp) to the director, are extracted from the numerical fits of the MSDs performed using a power law (See Eq. (1) in the main text) for the different particles. In the dilute range corresponding to the nematic phase formed by the host particles, both fdY21M and M13K07 rods exhibit a diffusive behavior over the whole time window with $\gamma_{\parallel,\perp} \approx 1$ in both the parallel and perpendicular directions, as shown in Figs. S2-right and S4. At higher concentrations, in the Smectic-A range close to Smectic-B phase, the slope of the log-log representation of the MSDs changes and suggests some anomalous diffusion. In this case, the exponent at short times (which is indicated in blue) is smaller than the one at longer times. This can be interpreted as an effect of crowding when approaching the crystalline phase, where voids are created in the adjacent smectic layers. This results in the release of the layer constraint of the initially confined particles, allowing them to jump.

For consistency of the results, we have therefore tuned the time range of the fit to maintain

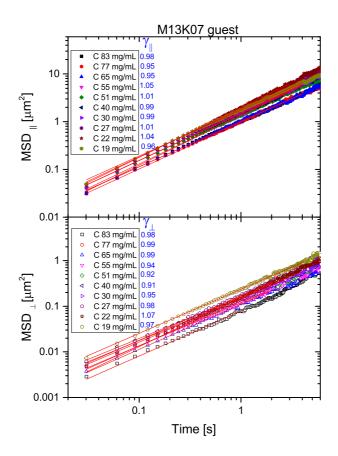


FIG. S4: Log-log representation of the parallel (top, solid symbols) and perpendicular (bottom, open symbols) mean square displacements (MSDs) for long guest M13K07 particles in the nematic range of the host particles. Blue values indicate the anisotropic exponents γ obtained from a power law fit performed according to Eq. (1) in the main text. They show a diffusive behavior both in parallel and perpendicular directions.

a high fit quality with a minimum value for the reduced chi-square.

IV. MOVIES

Time-stream DIC/fluorescence overlaid movies displaying the dual labeling of tracer amount of the host fdY21M (green) and guest M13K07 (red) particles in the smectic phase

 $(C_{fdY21M} = 93 \text{ and } 95 \text{ mg/mL}, \text{ respectively}).$ Scale bars indicate 3 μm .

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